

Spatial Mapping of Lipids and Elements by Mass Microscopy and Integration with LA-ICP-MS in the Diabetic Mouse Pancreata

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Introduction.

Pancreas endocrine function critically relies on the islets of Langerhans that secrete insulin and glucagon as well as other hormones. Previous pancreas LC-MS/MS lipidomics studies required islets isolation by a lengthy process involving tissue digestion and manual selection, potentially compromising metabolite basal levels [1]. MALDI-TOF imaging allows for in situ global assessment of lipids in pancreata cryosections. Our methodology was applied to a mouse model of diabetes, the adipocyte-specific-doxycycline-inducible mitochondrial ferritin (FtMT) overexpression, which displays massive beta-cell hyperplasia as its most striking phenotype [2].

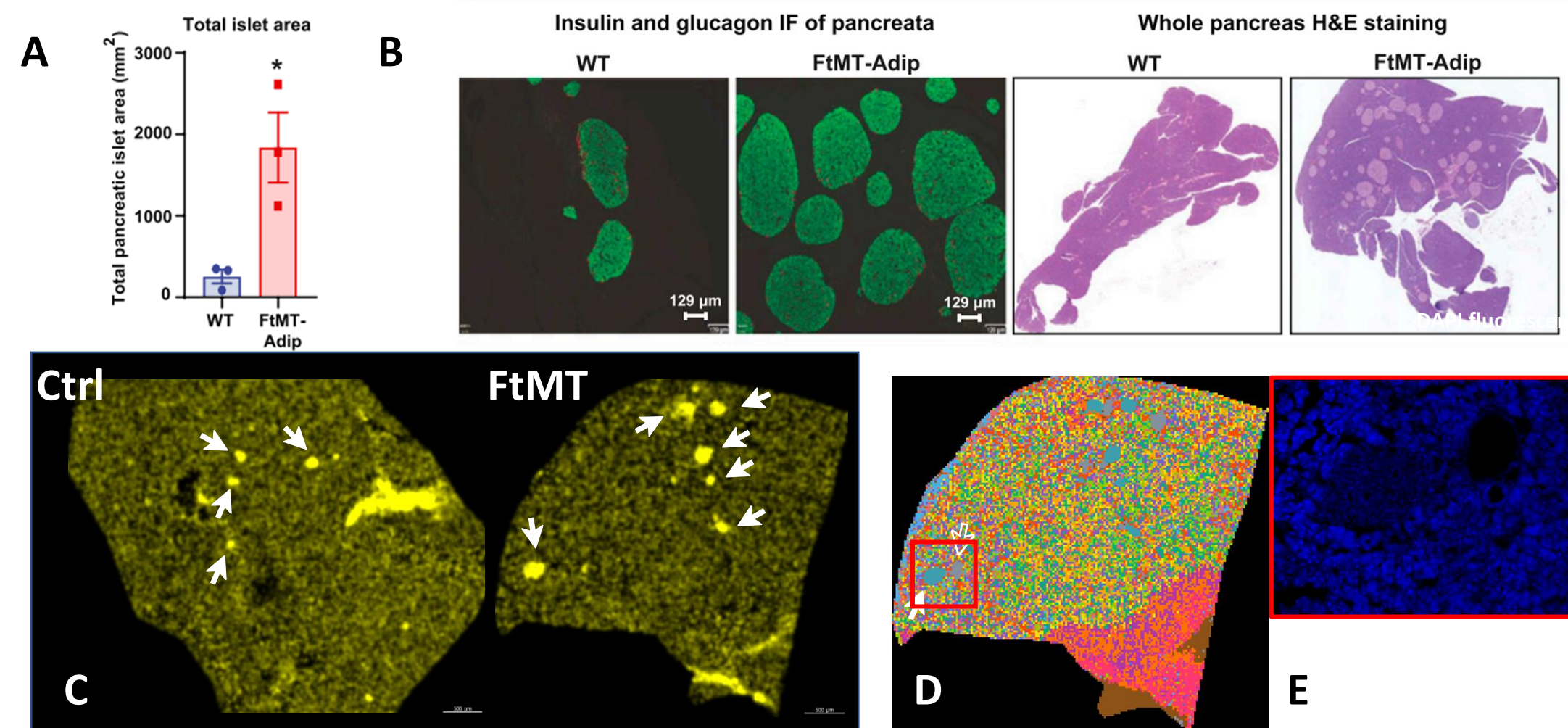


Figure 1. **A)** Total pancreatic islet area (mm²) [2]. **B)** Insulin IF staining of pancreata and (left) and H&E staining of whole pancreas from WT and FtMT-Adip mice [2]. **C)** Mouse pancreas false color mass spectrometry imaging (25 μ m spatial resolution) of SM 34:1;O₂ m/z 741.527 [M+K]⁺. Transgenic mouse pancreas displays considerably larger size islets. **D)** Unsupervised spatial segmentation calculation of MSI data (15 clusters) allows for differentiations of islets and blood vessel regions. **E)** DAPI fluorescence staining of red marked area in **D**.



Methods and Instrumentation.

Frozen mouse pancreata was cryosectioned into 10 μ m-thick sections. Tissue micro-sections were mounted on In₂O₃-SnO₂ (Millipore-Sigma, Burlington, MA). Automated matrix deposition system was achieved with an iMLayer™ (Shimadzu Corporation, Japan). 2-mercaptobenzothiazole (MBT, positive ion mode) and 1,5-diaminonaphthalene (DAN, negative ion mode) were used as MALDI matrix reagents. Mass spectrometry imaging (MSI) analysis was performed on an iMScope™ QT mass microscope (Shimadzu Corporation). Spatial elemental analysis was performed on an imageBIO266 (Elemental Scientific Lasers, USA) coupled to an ICPMS-2030 (Shimadzu Corporation). Imaging data processing was performed using IMAGEREVEAL™ MS software package (Shimadzu Corporation).

References. 1. Ye et al. J clin Invest 2018, 128(3):1178-1189 2. Kusminski et al. Diabetes 2020 69(3):313-330 3. MetaboAnalyst 5.0 www.metaboanalyst.ca

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Results.

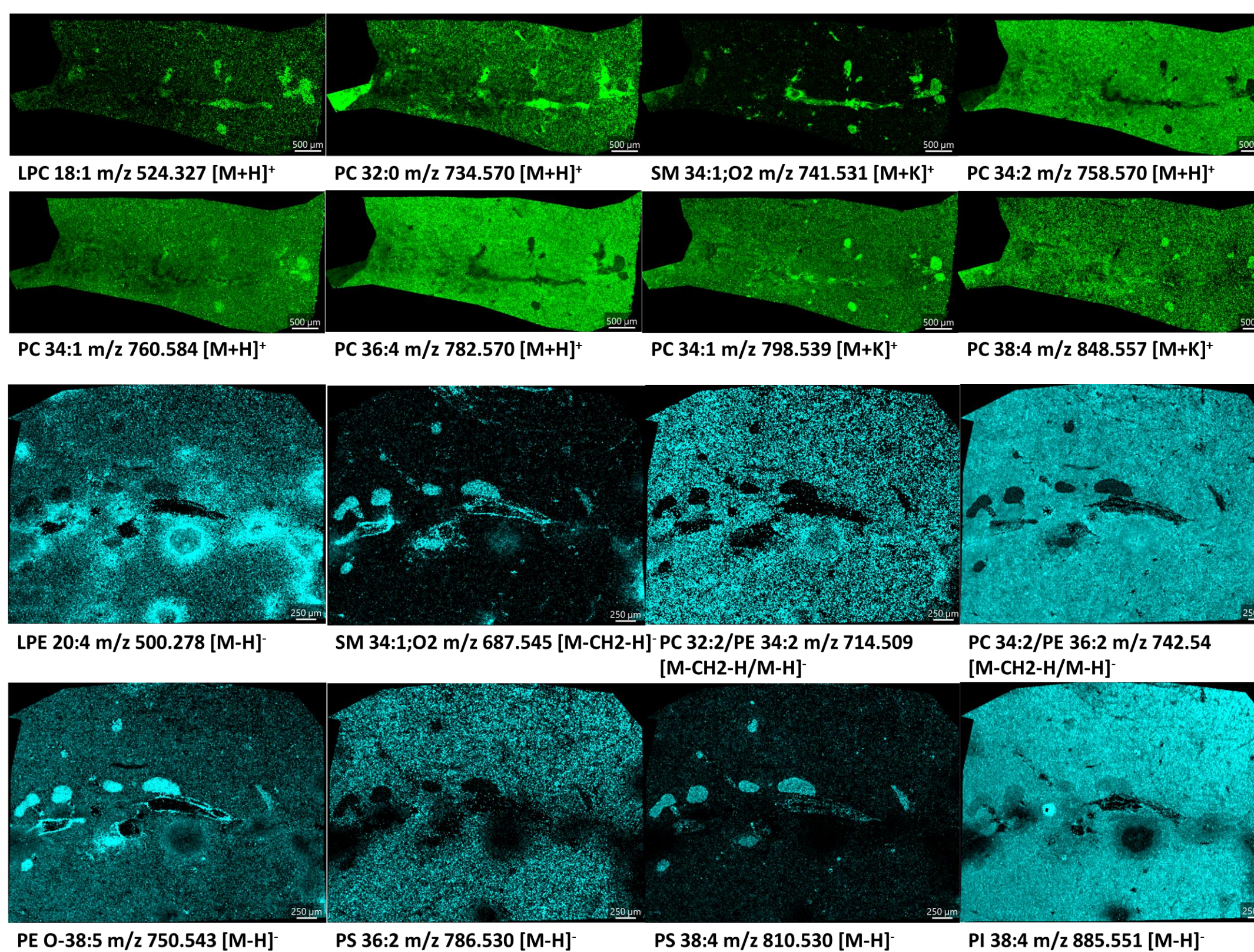


Figure 2. Top panel (green) positive ion mode analysis of wild-type (WT) pancreas cryosection at 6 μ m spatial resolution. Bottom panel (cyan) negative ion mode analysis of WT pancreas cryosection at 5 μ m spatial resolution.

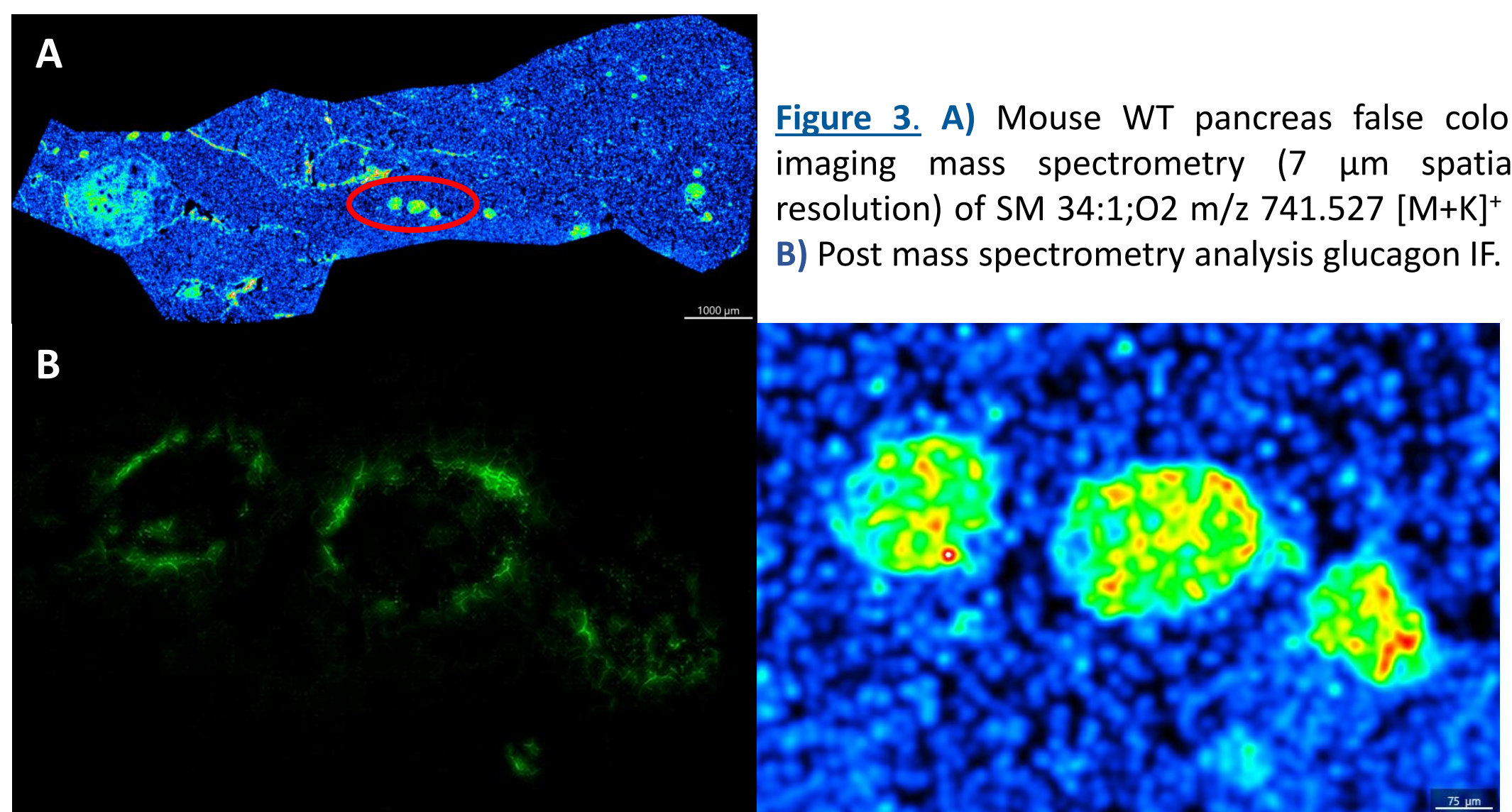


Figure 3. **A)** Mouse WT pancreas false color imaging mass spectrometry (7 μ m spatial resolution) of SM 34:1;O₂ m/z 741.527 [M+K]⁺. **B)** Post mass spectrometry analysis glucagon IF.

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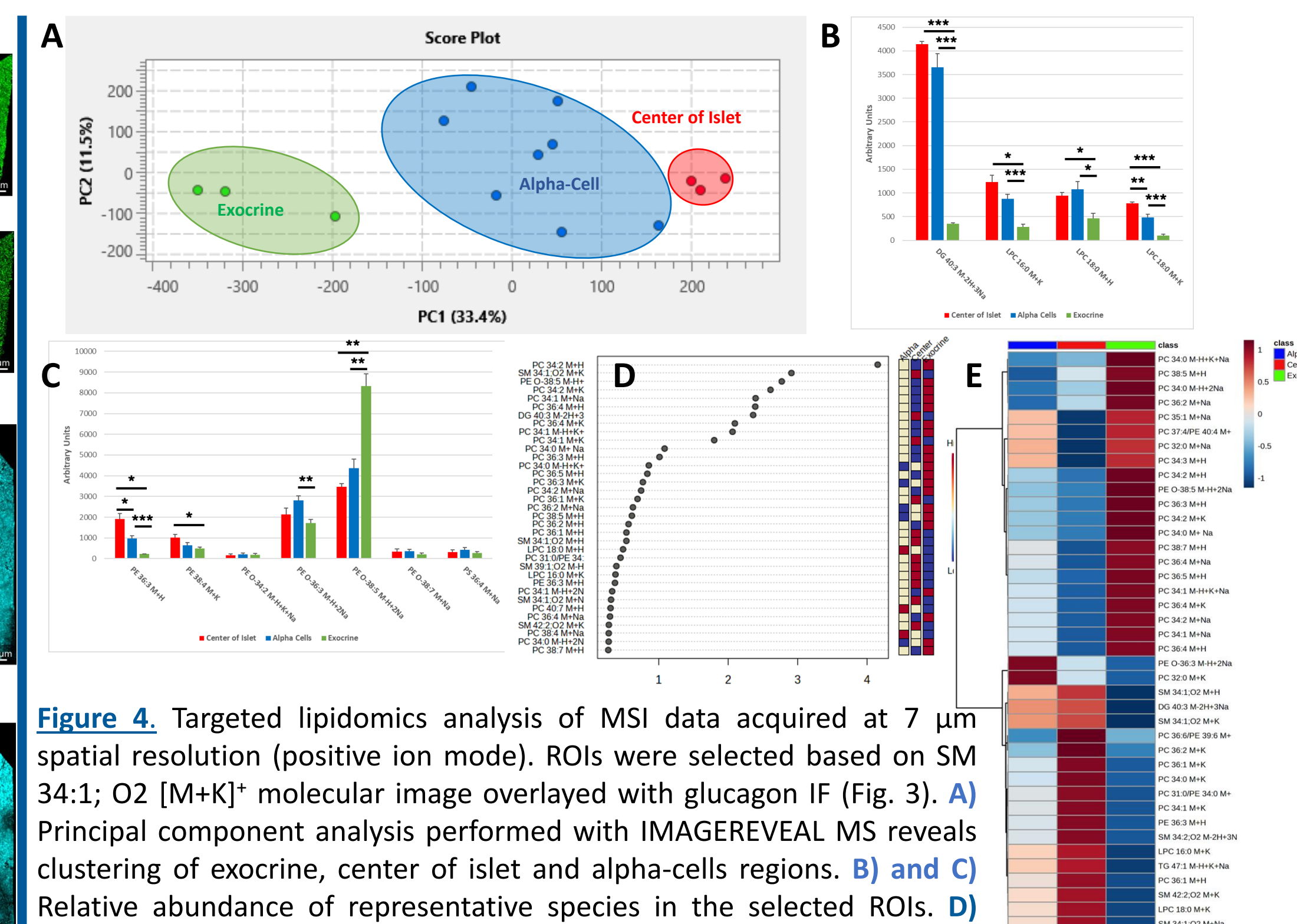


Figure 4. Targeted lipidomics analysis of MSI data acquired at 7 μ m spatial resolution (positive ion mode). ROIs were selected based on SM 34:1; O₂ [M+K]⁺ molecular image overlaid with glucagon IF (Fig. 3). **A)** Principal component analysis performed with IMAGEREVEAL MS reveals clustering of exocrine, center of islet and alpha-cells regions. **B) and C)** Relative abundance of representative species in the selected ROIs. **D)** Variable importance in projection (VIP) analysis. The top 35 metabolites with relative higher VIP score are shown [3] **E)** Heat map analysis of pancreata MSI lipidomics only. The top 40 most relevant features are shown [3].

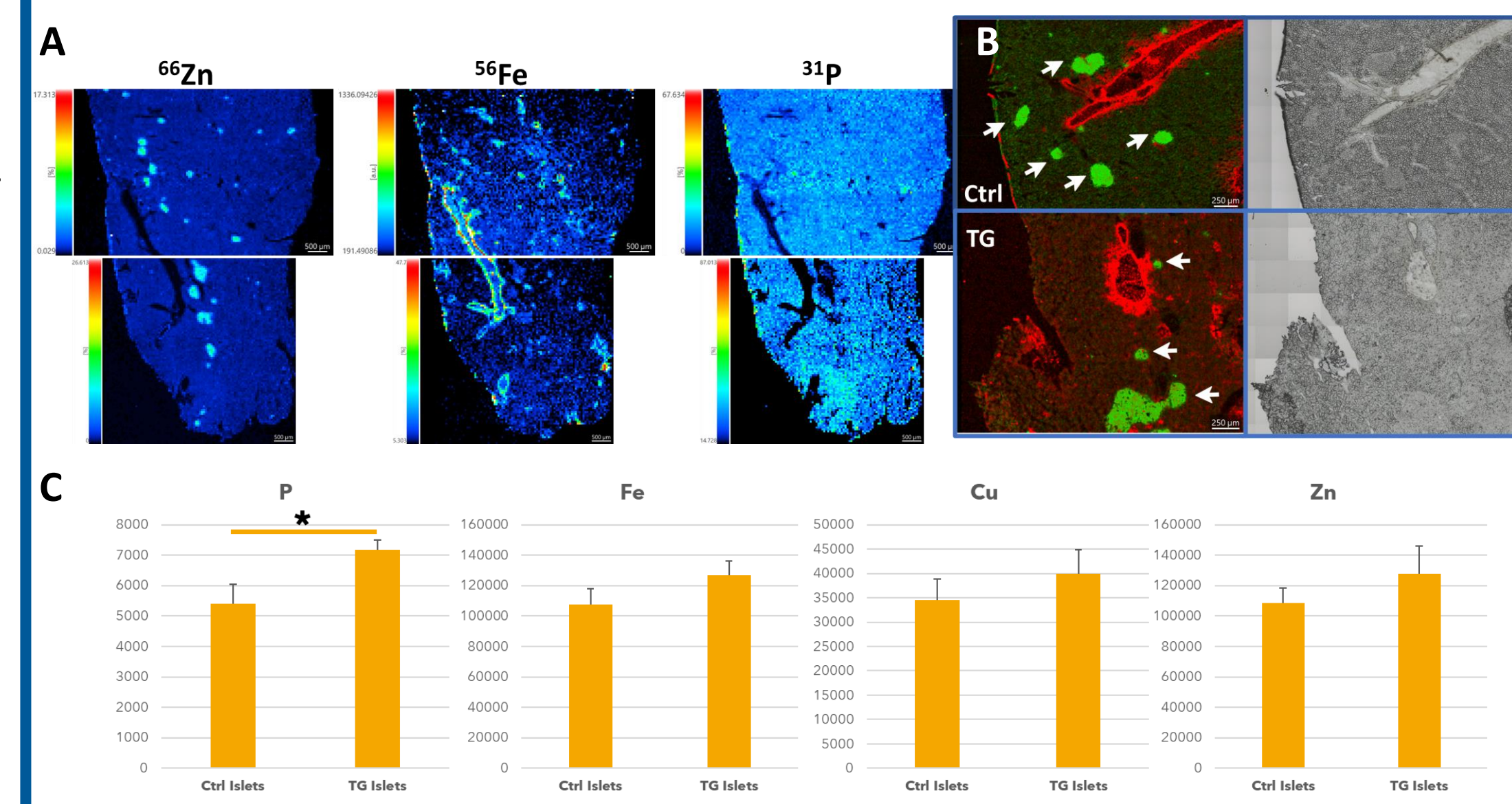


Figure 5. **A)** LA-ICP-MS imaging analysis of mouse pancreas at 50 μ m spatial resolution. Zn accumulates in the islets of Langerhans, Fe in blood vessels, while P is ubiquitous. **B)** Overlay of elemental images of Zn (green) and Fe (red) in control and FtMT overexpressing pancreas (analysis at 5 μ m spatial resolution). TG mice cryosection displays islets hyperplasia. High resolution optical images are also displayed. **C)** Elements relative abundance in islets and exocrine regions.