

Application note

Automated determination of THC contents in human urine

Category Sample preparation, solid phase extraction

Matrix Human urine

Method automated SPE, HPLC-MS

Keywords Drugs, THC, cannabis, Freestyle,

Automation, solid phase extraction



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Application ID



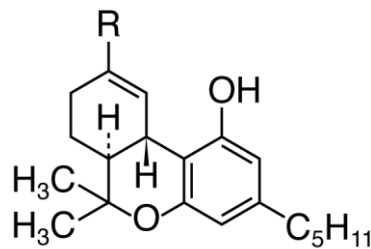
Abstract

This application note describes the automated determination of THC metabolites and its derivatives from human urine in the lower ng scale. It is shown, how the analysis can be automated using the automated sample preparation system FREESTYLE™ SPE from LCTech, Dorfen, Germany in combination with CHROMABOND® HR-X SPE cartridges. The subsequent analysis was done via HPLC-MS using a NUCLEOSHELL® RP 18 column.

Cannabis, also known as Marihuana or Hashish, is the most widely consumed drug in the world. Its consumption leads to mood-altering behavior like euphoria, relaxation and altered-time perception. A routinely consumption can lead to dependence and tolerance^{1,2}. In recent years there is also an increasing interest in therapeutic effects of cannabinoids and the development of potential cannabinoid medications. Therefore it is investigated for treatment of chronic pain, muscle spasticity, nausea and AIDS wasting disease, for example^{1,3,4}. This also leads to an increasing demand for the development of accurate and sensitive analytical methods for the quantification of cannabinoids in biological fluids¹. The measurement of urine cannabinoids is necessary for pharmacokinetic studies, drug treatment, workplace drug testing and for drug impaired driving investigations. Detailed urine collection procedures with comprehensive chain-of-custody documentation have been developed for forensic applications¹.

In this application note the analysis of Δ^9 -tetrahydrocannabinol (THC), the major psychoactive component of cannabis, and its derivatives 11-Hydroxy- Δ^9 -tetrahydrocannabinol (THC-OH) and 11-Nor- Δ^9 -tetrahydrocannabinol COOH (THC-COOH) from human urine is shown. The time limiting factor for the lab staff, especially for high-throughput laboratories is the solid-phase extraction prior to the analysis, as all steps have to be done manually. In order to automate the whole process, an automated sample-preparation system was used, the FREESTYLE™ SPE system from LCTech, Dorfen, Germany. This system is a modular and completely automated sample preparation system that can work completely unsupervised. Every manual SPE method can be fast and easily transferred to the FREESTYLE™ system. In combination with CHROMABOND® HR-X columns a completely automated sample preparation with excellent recovery rates was established for the analysis of THC and its derivatives from human urine. The subsequent analysis was done using HPLC-MS with a MACHEREY-NAGEL NUCLEOSHELL® RP 18 column for fast and excellent results.

Figure 1 shows the molecular structures of THC and its derivatives THC-OH and THC-COOH.



Analyte	-R	Formula	Mass (g/mol]
THC	-CH ₃	C ₂₁ H ₃₀ O ₂	314,5
THC-OH	-CH ₂ -OH	C ₂₁ H ₂₈ O ₄	330,5
THC-COOH	-COOH	C ₂₁ H ₃₀ O ₃	344,4

Figure 1:
Molecular structures of THC and its derivatives THC-OH and THC-COOH



Materials

All used solvents and chemicals are of p.A. grade ($\geq 99.0\%$), all solvents used for the HPLC-MS analysis are of ULC/MS grade.

Experimental sample preparation

Samples of human urine are spiked with a standard solution to get defined concentrations of THC, THC-OH and THC-COOH. Therefore the standard solutions of THC, THC-OH and THC-COOH are combined to get a standard solution that contains all of the three analytes in defined concentration.

5 mL urine are spiked with 50 μL of that combined standard solution.

All measurements were conducted in triple determination.

Table 1: Overview of the standard solutions and spiked urine sample

Solution	Concentration
Δ^9 -Tetrahydrocannabinol from Lipomed THC-135-1LE (THC)	(10 $\mu\text{g}/\text{mL}$ in methanol)
11-Hydroxy- Δ^9 -tetrahydrocannabinol from Lipomed THC-318-1LM (THC-OH)	(10 $\mu\text{g}/\text{mL}$ in methanol)
11-Nor- Δ^9 -tetrahydrocannabinol COOH from Lipomed THC-316-1LM (THC-COOH)	(10 $\mu\text{g}/\text{mL}$ in methanol)
Combined standard solution	(1×10^3 mg/mL (each))
Spiked urine sample	10 ng/mL (5 mL urine spiked with 50 μL standard solution)

As THC and its metabolites are secreted as glucuronide conjugates in urine, it is necessary to cleave the glucuronide moiety before the solid phase extraction is conducted. In this case the hydrolysis was done via base to avoid the formation of glucuronides in the spiked urine samples.

300 μL NaOH solution (10 M) were given to 5 mL of the urine samples and shaken for 15 min at 60°C in a heating block. The solution is cooled to room temperature and 200 μL acetic acid are added. At last, 2 mL ammonium acetate solution (50 mM) is given to the solution and the pH is adjusted to pH 6-7 with acetic acid or NaOH solution.

Solid phase extraction (manual)

Before adapting the solid phase extraction to the automated FREESTYLE™ SPE system, a manual method for the extraction was developed. The used conditions are given in Table 2.



Table 2: Conditions of the SPE method (manual)

Column	CHROMABOND® HR-X, 3 mL, 200 mg
Column conditioning	2 mL methanol (2 mL/min) 2 mL H ₂ O (Millipore) (2 mL/min) 2 mL ammonium acetate buffer (50 mM) (2 mL/min)
Loading	The sample is applied onto the column with a speed of 1-2 mL/min
Washing step	5 mL methanol - H ₂ O (Millipore) (V:V, 3:7) Drying for 10 min
Elution step	3 mL hexane - ethyl acetate - glacial acetic acid (V:V:V, 75:25:1)

Solid phase extraction (automated)

The manual solid phase extraction can be easily transferred to the FREESTYLE™ SPE system for an automated sample preparation. The software of the FREESTYLE™ SPE system contains pre-defined method wizards for every step making the method development very fast and easy. Up to fifteen solvents can be connected to the system.



Figure 2: Predefined steps for the automated SPE method



Figure 3 shows the method wizard for the sample load step as example. For every step it can be defined which port of solvent is used, which volume of solvent is used and the speed the solvents is forced through the cartridge.

Load

Transfer Type: Quantitative transfer over sample loop

Input Vial Type: Type1@16

Sample Volume: 13 ml

Suction Speed: 10 ml / min

Dispensing Speed: 1 ml / min.

Waiting time after dosage: 25 s

Waiting Time after Step: 150 s

Wash-down volume: 1 ml

Repetitions: 3 x

Suction Speed: 1.1 ml / min

Solvent from Port: Port9: water +methanol,

Tube rinse volume: 1 ml

Dispensing Speed: 0.3 ml / min

Dispense: into Waste Online - EVA into vial

Don't open seal of SPE column while loading

34.8 min

Figure 3: Software wizard for the loading step of the automated SPE method

Table 3 gives an overview over the conditions used for every step of the automated SPE method.

Table 2: Conditions of the SPE method (automated)

Column	CHROMABOND® HR-X, 3 mL, 200 mg
Column conditioning	2 mL methanol (2 mL/min) 2 mL H ₂ O (Millipore) (2 mL/min) 2 mL ammonium acetate buffer (50 mM) (2 mL/min)
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Eluent exchange

In order to ensure that the analytes are dissolved in an HPLC/MS compatible solvent, an eluent exchange has to be performed prior to the analysis. Therefore, the eluates from the solid phase extraction are evaporated to dryness at 40°C under nitrogen. The residue is dissolved in 1 mL methanol.



HPLC-MS/MS analysis

The subsequent HPLC-MS analysis was performed on a Dionex Ultimate 3200 HPLC system in HPG configuration with an ABSciex 3200 MS detector. The following conditions were used.

Table 4: Method for the HPLC-MS analysis

Column	NUCLEOSHELL® RP 18, 2,7 µm, 50 x 2 mm		
Eluent A	H₂O (Millipore) + 0.1% formic acid		
Eluent B	acetonitrile + 0.1% formic acid		
Gradient	Time [min]	%A	%B
	0.0	50	50
	2.5	0	100
	5.0	0	100
	5.1	50	50
	7.5	50	50
Flow rate	0.3 mL/min		
Injection Volume	5 µL		
Column Temperature	40 °C		
Detection	API 3200 Ion source: ESI Ionisation mode: positive Curtain gas: 20 psig Ionspray Voltage: 5500 V Temperature: 550 °C Gas 1: 20 psig Gas 2: 20 psig CAD: 6.0 psig MRM-Transitions		
Run Time:	7.5 min		



Results

Two mass-transitions per analyte were detected as qualifier and quantifier. Figure 4 shows the two transitions for the THC-OH as example.

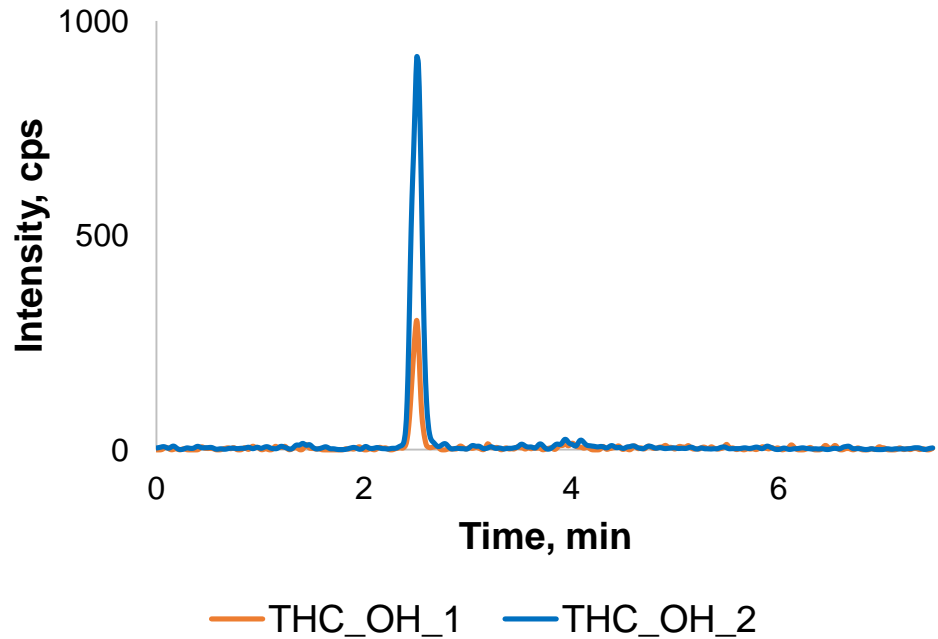


Figure 4: Chromatogram of the HPLC-MS analysis

Table 5 shows the results of the MS analysis.

Table 5: Results of the MS analysis

Analyte	[M+H] ⁺	Q1 (Quantifier)	Q2 (Qualifier)
THC	315.2	193.2	123.1
THC-OH	331.2	313.3	43.1
THC-COOH	345.2	327.3	299.4

Based on a calibration with different standard solutions, the peak areas were integrated and the recovery rates for both, the manual and the automated SPE method were calculated.

In Table 6 the recovery rates as results of a triple determination of both methods are compared to each other. It can be seen, that for the automated method even higher recovery rates than for the manual method are found.



Table 6: Recovery rates for the manual and automated SPE methods (average from triple determination)

Analyte	Recovery rate [%]	
	manual	FREESTYLE™ SPE
THC	80 ± 3	90 ± 9
THC-OH	77 ± 10	80 ± 6
THC-COOH	98 ± 8	93 ± 8

Conclusion

Using the automated FREESTYLE™ SPE sample preparation system in combination with CHROMABOND® HR-X cartridges and a subsequent LC-MS analysis using NUCLEOSHELL® RP 18 showed reliable results for the analysis of THC and its metabolites from human urine. Using the automated system even better recovery rates were found than using a manual SPE method.

Literature

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Physical properties of recommended columns

Table 7: Properties of the recommended SPE cartridges



For the solid phase extraction, CHROMABOND® HR-X SPE cartridges from MACHEREY-NAGEL are used. Table 7 gives an overview over the physical properties of those cartridges.

Stationary Phase	CHROMABOND® HR-X
Carrier material	Polymeric
Surface modification	none
Pore size	55-60 Å
Surface area (BET)	1000 m ² /g
Particle size	85 µm
Shape	Spherical
Cartridge size, amount of sorbens	3 mL, 100 mg
REF number (MN)	730931 (30 pc./pg.)

For the HPLC-MS analysis a NUCLEOSHELL® RP 18 column is recommended. In Table 8 the properties of the used column are summarized.

Table 8: Properties of the recommended HPLC column



Stationary phase	NUCLEOSHELL® RP 18
USP Code	L1
Pore size	90 Å
Surface area	130 m ² /g
%C	7.8
Endcapping	Multi-endcapping
Particle size	2.7 µm
Shape	Spherical
Dimensions	50 x 2 mm
REF Number (MN)	763132.20



Recommended Instrumentation

The automated solid-phase extraction is performed on a FREESTYLE SPE system from LCTech, Dorfen, Germany. Its modular configuration allows to set up the system tailor-made to special desires. Table 9 summarizes the modules used for this application note.

Table 9: Used modules of the FREESTYLE™ system

FREESTYLE™ Module	REF number (LCTech)
FREESTYLE™ Basic	12663
FREESTYLE™ SPE	12668
Upgrade to use 6 instead of 3 solvents	12952
Discharge function for nitrogen	12905
Rack for 60 x 4 mL sample flasks	11926
Rack for 30 x 16 mL sample flasks	11933
Rack for up to 18 SPE columns	13416
Carrier for racks, 100 mm broad	11915
Carrier for racks, 120 mm broad	12103
Lock ring for 3 mL standard SPE columns (10 pc./pg.)	12334
Plug for 3 mL SPE columns (100 pc./pg.)	13491
Screw vial, about 16 mL (100 pc./pg.)	V0016
Plug with hole (100 pc./pg.)	V0016-SL
Septa for 16 mL vials (100 pc./pg.)	V0016-D

For more information about the system click here or contact LCTech:

<http://www.lctech.de/en/automated-preparation/freestyle-spe>



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