

Solid Phase Micro Extraction of Tea Flavor Components

Application Note

Food and Flavor

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Abstract

Tea flavors can vary from spicy to flowery to fruity and any combination thereof. Moreover, the flavor profile of tea can depend on where the tea leaves are grown, the brewing time and temperature, the processing of the tea leaves, and the type of leaf used. Using Head Space Solid Phase Micro Extraction (HS-SPME) sampling in conjunction with Gas Chromatography/Mass Spectrometry (GC/MS) for separation and analysis, assorted teas will be examined for their varied flavor components.

Introduction:

Green, Black and Jasmine Teas are all made from the same plant. They derive their flavor distinctions from how they are processed. Green tea leaves experience very little treatment while black tea leaves are oxidized before they are dried. This oxidation process causes the leaves to darken and produces a much stronger flavor. Jasmine tea, on the other hand, is green tea leaves scented with jasmine flowers.

In order to determine the differences in the three teas, the aroma compounds were extracted from the headspace of the brewed tea using Solid Phase Micro Extraction (SPME). The SPME fiber was then desorbed onto a column in a Gas Chromatograph (GC) for separation and analyzed using a Mass Spectrometer (MS). The resulting chromatograms were then analyzed in order to determine the flavor compounds of each respective tea and how they differed from one another.

Experimental:

The EST Analytical FLEX autosampler was used in order to automate the sampling process. A Divinylbenzene Carboxen/Polydimethylsiloxane (DVB/CAR/PDMS) SPME fiber with a 50/30 μ m film thickness was found to be the most efficient SPME fiber for this examination. The Shimadzu QP2010 SE GCMS was fitted with a SPME liner and a Restek Rxi-5 Sil MS column was used for separation and analysis.

Headspace SPME is a non-exhaustive sampling technique so the experimental conditions had to be optimized in order to make the extraction technique both efficient and reproducible. The FLEX suite software simplified the sample method development process with the ease of its drag and drop method builder. The autosampler and GCMS experimental conditions developed for this analysis are listed in Tables 1 and 2.

| Autosampler | | FLEX |
|--------------------------|--|-----------|
| General | | |
| Method Type | | SPME |
| GC Ready | | Continue |
| GC Cycle Time | | 30min |
| Constant Heat Mode | | Yes |
| Sample Incubate Agitate | | |
| Incubation Temp. | | 80°C |
| Incubation Time | | 0.2min |
| Agitation Duration | | 0.0min |
| Extraction | | |
| Fiber Guide Depth | | 45% |
| Sample Vial Fiber Depth | | 1cm |
| Extraction Time | | 10.1min |
| Fiber Extraction Agitate | | Yes |
| Agitate Type | | Oscillate |
| Agitate Duration | | 10.0min |
| Wait | | |
| Wait on Input | | Yes |
| Wait Input | | GC Ready |
| Desorption | | |
| Injection Port | | A |
| Fiber Guide Speed | | 40% |
| Fiber Guide Depth | | 50% |
| Fiber Insertion Speed | | 75% |
| Fiber Insertion Depth | | 1cm |
| Fiber Desorption Time | | 2.0min |
| Injection Start Output | | Start |
| Condition Fiber | | |
| Fiber Temp | | 250°C |
| Condition Time | | 5.0min |
| Fiber Guide Speed | | 60% |
| Fiber Guide Depth | | 60% |
| Fiber Insertion Speed | | 20% |
| Fiber Insertion Depth | | 1cm |

Table 1: FLEX Autosampler Experimental Parameters

| GC/MS | | Shimadzu QP 2010 SE |
|--------------------------|--|---|
| Inlet | | Split/Splitless |
| Inlet Temp. | | 250°C |
| Inlet Head Pressure | | 51.6kPa |
| Mode | | Splitless |
| Injection Pulse Pressure | | 100kPa for 2.0 min |
| Carrier Gas Split Ratio | | 10:1 |
| Desorption | | 2.0min at 250 °C |
| Column | | Rxi-5 Sil MS 30.0m X 0.25mm X 0.25µm |
| Oven Temp. Program | | 45°C hold for 1.0 min., ramp 20°C/min to 275°C, hold for 1.5min, 14min run time |
| Column Flow Rate | | 1.0ml/min |
| Gas | | Helium |
| Linear Velocity | | 36.2ml/min |
| Source Temp. | | 220°C |
| MS Transfer Line Temp. | | 220°C |
| Scan Range | | m/z 35-500 |
| Event Time | | 0.20sec |
| Solvent Delay | | 2.1min |

Table 2: GC/MS Experimental Parameters

The tea leaves were acquired in China and each tea was prepared in the same manner. Two grams of the tea leaves were placed in 250 milliliters of de-ionized water at 80°C. The tea was allowed to steep in the heated water for five minutes. Ten milliliters of the prepared tea was then placed into a prepared 20 milliliter headspace vial. The headspace vials each had one half gram of sodium chloride in them in order to aid in analyte extraction. Five replicates of each tea were sampled and analyzed so as to verify reproducibility.

The results were analyzed in order to compare the relative response of the flavor compounds that the individual teas had in common. Table 3 and Figures 1 and 2 display the results of this analysis. Figures 3 through 5 presents labeled chromatograms of each tea in order to demonstrate the differences in the overall analyte response of each tea.

| Compound | Odor | Black Tea | Green Tea | Jasmine Tea |
|----------------------------------|-----------------------|-----------|-----------|-------------|
| hexanal | grass | 2861326 | 5433817 | |
| 2-amino-6-methyl-benzoic acid | fruity, musty | 4713330 | 2439315 | 3121718 |
| benzaldehyde | fruity, almond cherry | 1914082 | 98979 | 2726334 |
| beta-myrcene | spicy | 7401212 | | 6131413 |
| d-limonene | citrus | 3261019 | | 1666963 |
| geraniol | floral | 3049259 | 1537017 | 3952291 |
| methyl salicylate | minty | 6103515 | | 41879957 |
| methyl anthranilate | fruity | 2389211 | | 31610862 |
| 2,6-di-tert-butyl p-benzoquinone | new mown hay | 6386643 | 4740502 | 4122720 |
| trans-beta ionone | floral, woody | 2983205 | | 2937920 |
| isopropyl myristate | faint oil | 1115581 | 2251202 | 1248786 |
| isopropyl palmitate | faint oil | 1527085 | 2247817 | 2386125 |
| linalool | citrus, floral | 72921687 | 3045770 | 166582273 |

Table 3: Flavor Compound Responses

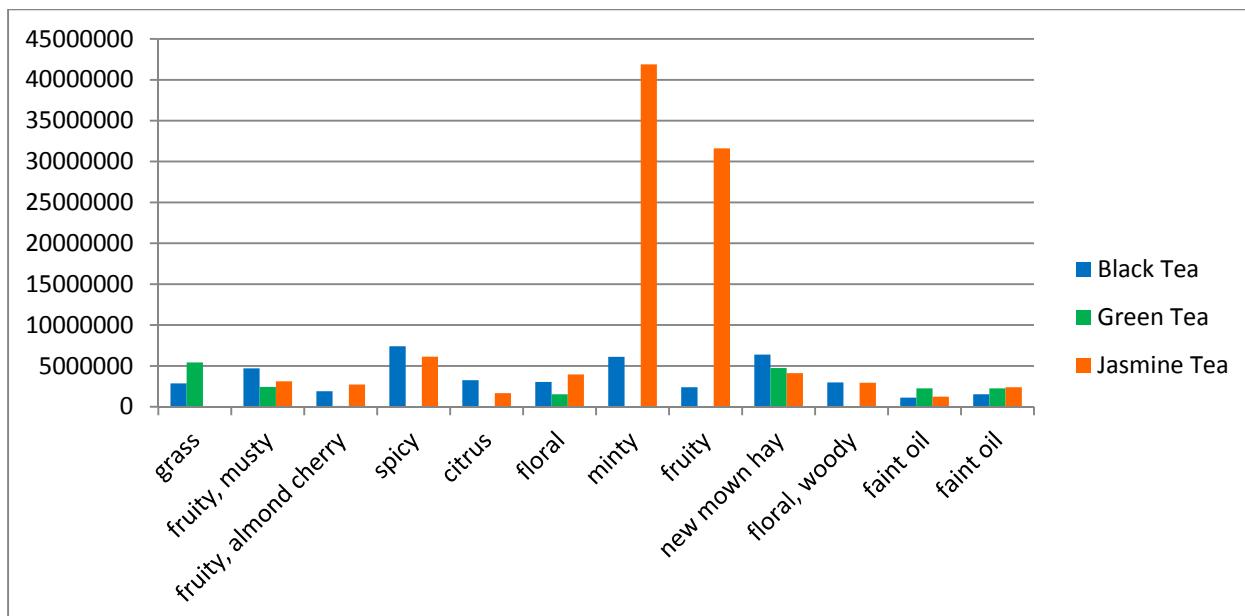


Figure 1: Diagram of Flavor Compound Comparison

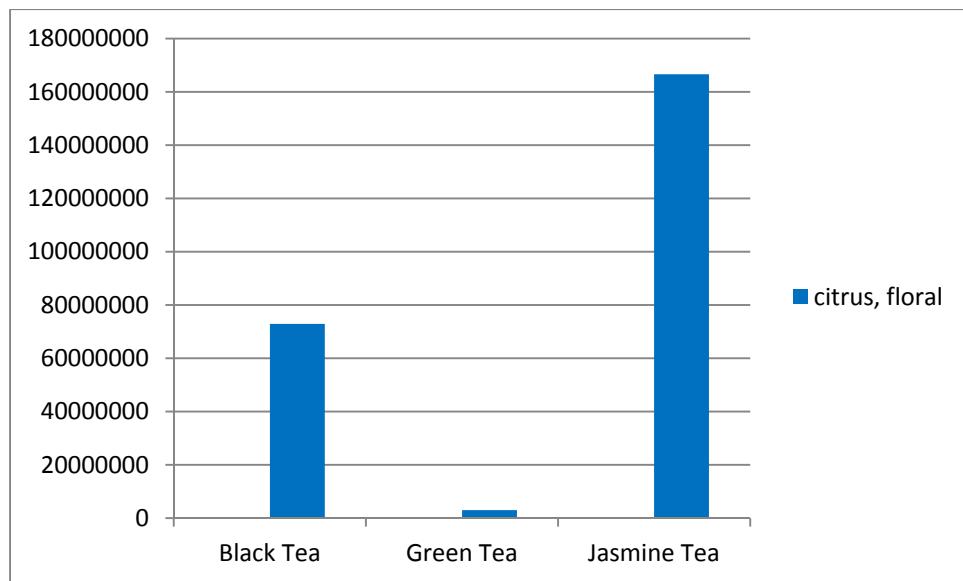


Figure 2: Difference in Linalool Response for Each Tea

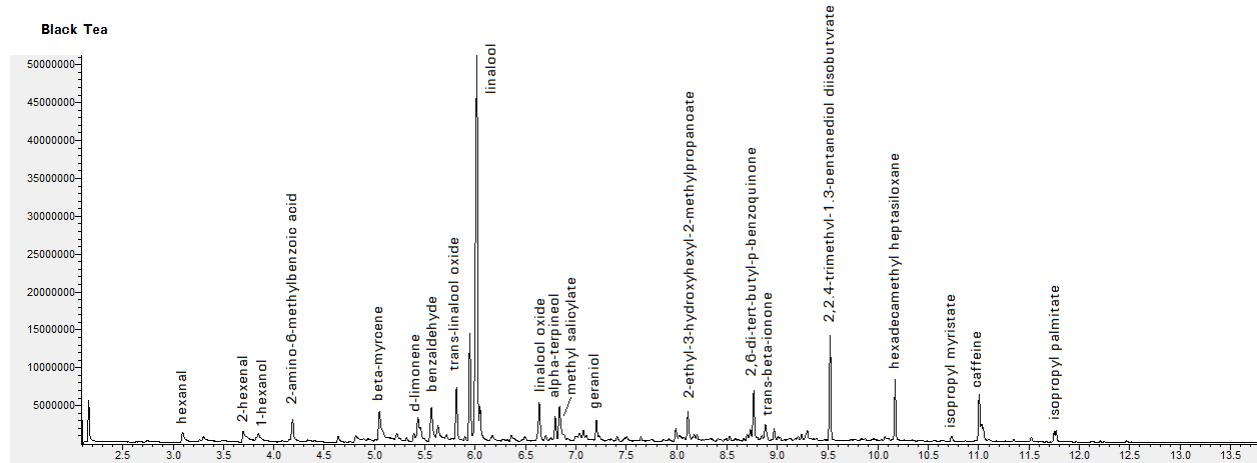


Figure 3: Chromatogram of Black Tea

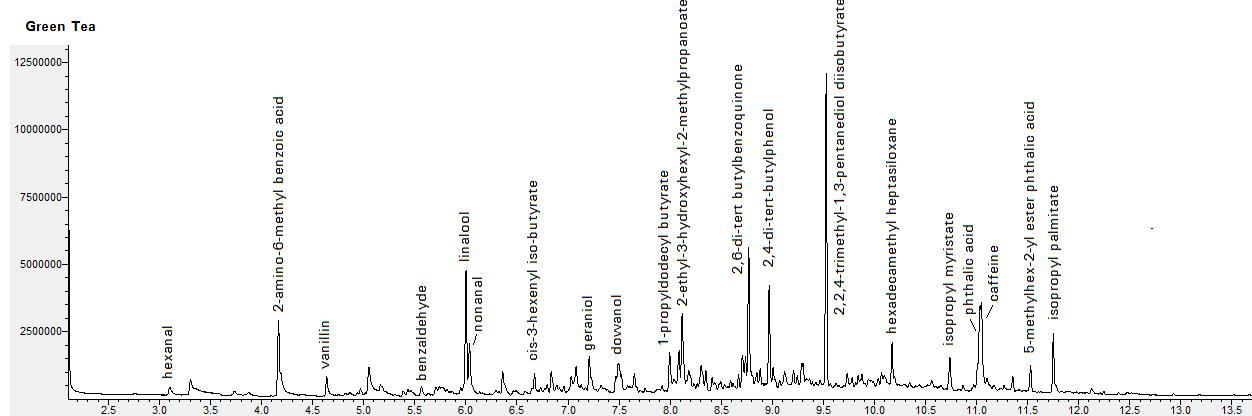


Figure 4: Chromatogram of Green Tea

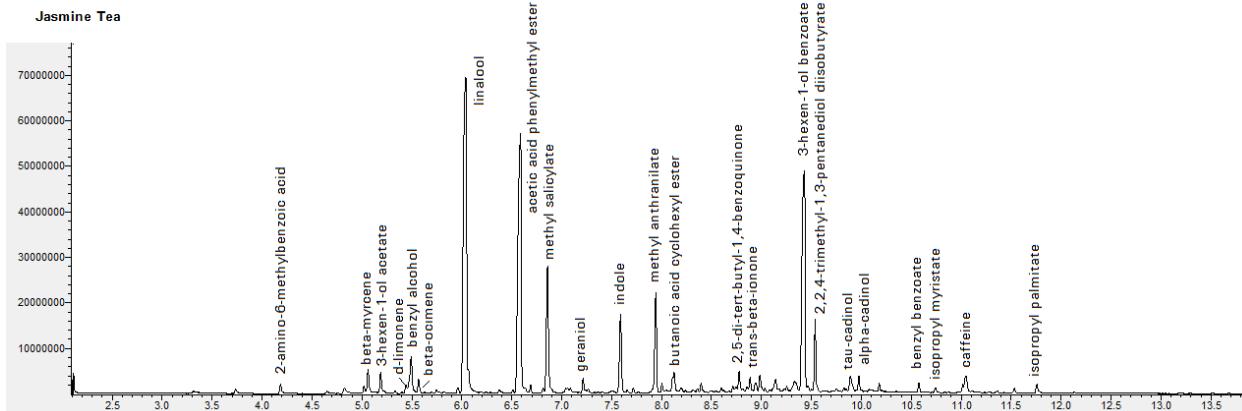


Figure 5: Chromatogram of Jasmine Tea

Conclusions:

Overall, the black and the jasmine teas had the most flavor compounds in common. Linalool, which gives tea a floral flavor, was common to all of the teas, however the jasmine tea had over twice the amount of linalool than the black tea and 50 times the amount in the green tea. The chromatography of each tea was quite unique from the others with green tea having the most complex chromatogram. The FLEX provided an excellent platform for the SPME sampling of the brewed teas. The FLEX method builder software enabled efficient method development and using the Shimadzu LabSolutions software the analysis of the flavor compounds in the tea was a straightforward process.

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