LC-MS/MS Method for the Determination of Steroids from Urine Using SOLA and Core Enhanced Technology Accucore HPLC Column.

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Key Words

- SOLA Cartridges and Plates
- Accucore
- Solid Core
- Sore Enhanced Technology
- Steroids
- Hydrocortisone
- Cortisone
- Corticosterone
- 11-α hydroxyprogesterone
- Betamethasone

Abstract

This application note demonstrates the use of the Thermo Scientific SOLA well plates for the extraction of steroids from urine and the subsequent fast separation using a Thermo Scientific Accucore RP-MS column.

Introduction

SOLATM products are a revolutionary new Solid Phase Extraction (SPE) product range. This first in class SPE product range introduces next-generation, innovative technological advancements, giving unparalleled performance characteristics compared to conventional SPE, phospholipid and protein precipitation products.

This includes:

- Higher levels of reproducibility
- · Higher levels of extract cleanliness
- Reduced solvent requirements
- Increased sensitivity

SOLA products have significant advantages for the analyst when processing compounds in complex matrices particularly in high throughput bioanalytical and clinical laboratories where reduced failure rate, higher analysis speed and lower sample/solvent requirements are critical.

The increased performance from SOLA products provides higher confidence in analytical results and lowers cost without compromising ease of use or requiring complex method development.

AccucoreTM HPLC columns use Core Enhanced Technology to facilitate fast and high efficiency separations. The 2.6 μm diameter particles are not totally porous, but rather have a solid core and a porous outer layer. The optimized phase bonding creates a series of high coverage, robust phases. Accucore RP-MS uses an optimized alkyl chain length for more effective coverage of the silica surface. This coverage results in a significant reduction in secondary interactions and thus highly efficient peaks with very low tailing. The tightly controlled 2.6 μm diameter of Accucore particles results in much lower backpressures than typically seen with sub-2 μm materials.

Hydrocortisone, cortisone, corticosterone and 11- α hydroxyprogesterone belong to the progesterone class of steroid horomones. Progesterones are characterized by the 21-carbon backbone and are primarily used to maintain pregnancy. The extraction of these steroids from urine, their separation on an Accucore RP-MS column and their quantification by LC-MS/MS is demonstrated in this application note.



Experimental Details

| Consumables | Part Number |
|--|-----------------------|
| Fisher Scientific LC-MS grade water | W/0112/17 |
| Fisher Scientific LC-MS grade acetonitrile | A/0638/17 |
| Fisher Scientific LC-MS grade formic acid | A117-50 |
| Hydrocortisone, cortisone, corticostereone, 11- α hydbetamethasone (IS) | lroxyprogesterone and |

| Sample Preparation | Part Number | |
|---------------------|---|---------------|
| Matrix: | urine | |
| Cartridge type: | SOLA 96 well plate | 60309-001 |
| Conditioning stage: | 1 mL methanol, 1 mL water | |
| Application stage: | 1 mL spiked urine containing internal standard for corticosterone and 11- α hydroxyprogesterone. | |
| | 1 mL urine for hydrocortisone | and cortisone |
| Washing stage: | 200 µL of 20:80 (v/v) methanol / water | |
| Elution stage: | 2 x 200 μL methanol | |
| Additional stage: | Evaporate to dryness under a gentle stream o nitrogen and reconstitute in 200 µL water | |



| Separation Conditions | | | Part Number | |
|--------------------------------|---|----|--------------------|--|
| Instrumentation: | Thermo Scientific Accela | | | |
| Column: | Accucore RP-MS 2.6 μm, 100 x 2.1 mm | | 17626-102130 | |
| Mobile phase: | Nobile phase: A: water + 0.1% formic acid | | | |
| | B: acetonitrile + 0.1% formic acid | | | |
| Gradient: | Time (minutes) | %B | | |
| | 0.0 | 25 | | |
| | 0.02 | 25 | | |
| | 4.00 | 75 | | |
| | 4.50 | 75 | | |
| | 4.51 | 25 | | |
| | 6.00 | 25 | | |
| Flow rate: | 0.6 mL/min | | | |
| Column temperature: | 25 °C | | | |
| Injection details: | 2.5 μL | | | |
| Weak Injection wash solvent: | 20:80 (v/v/v) acetonitrile / water | | | |
| Strong injection wash solvent: | tt: 45:45:10 (v/v) acetonitrile / acetone / isopropanol | | | |

MS Conditions

Instrumentation: Thermo Scientific TSQ Vantage

| Ionization conditions | HESI |
|---------------------------|------|
| Polarity | +ve |
| Spray voltage (V) | 3000 |
| Vaporizer temp (°C) | 300 |
| Sheath gas pressure (Arb) | 60 |
| Aux gas pressure (Arb) | 30 |
| Capillary temp (°C) | 300 |
| Collision pressure(mTorr) | 1.5 |
| Scan time (s) | 0.02 |
| Q1 (FWHM) | 0.7 |
| Q3 (FWHM) | 0.7 |

Table 1. Vantage[™] conditions

| Compound | Hydro- cortisone | Cortisone | Corti- costerone | 11-a hydroxypro -gesterone | Beta- methasone (IS) |
|--------------------------|---------------------|-----------|---------------------|----------------------------------|----------------------------|
| Precursor (m/z) | 363.23 | 361.30 | 347.29 | 331.29 | 393.28 |
| Product (m/z) | 121.03 | 163.12 | 329.27 | 295.24 | 373.26 |
| Collision energy (eV) | 21 | 20 | 10 | 11 | 5 |
| S-lens (RF voltage) | 87 | 102 | 82 | 84 | 63 |

Table 2. Transition details

Data Processing

| Software: Thermo Scientific LC QUAN | |
|-------------------------------------|--|
|-------------------------------------|--|

Results

As shown in Figure 1, the four steroids were separated in less than 3 minutes using an Accucore RP-MS column.

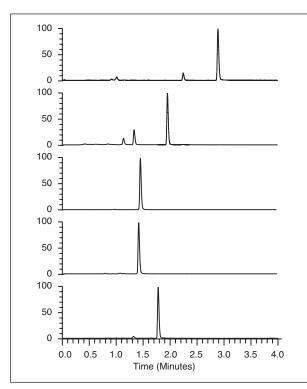


Figure 1. Extracted ion chromatograms of 11- α hydroxyprogesterone (1), corticosterone (2), cortisone (3), hydrocortisone (4) and internal standard betamethasone (5) from urine using SOLA well plate

Hydrocortisone results

Hydrocortisone standards prepared in water were linear over the dynamic range of 1 to 1000 ng/mL with an r² of 0.9937 (Figure 2). As shown in Table 3, six replicates of urine were extracted using the SOLA 96 well plates and the endogenous level of hydrocortisone was found to be 17.3 ng/mL with precision of 11.1%.

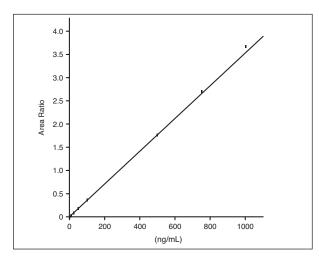


Figure 2. Linear dynamic range of 1 to 1000 ng/mL with an $\rm r^2$ of 0.9937 for hydrocortisone standards

Cortisone results

Cortisone standards prepared in water were linear over the dynamic range of 1 to 1000 ng/mL with an r² of 0.9981 (Figure 3). As shown in Table 3, six replicates of urine were extracted using the SOLA well plates and the endogenous level of cortisone was found to be 55.6 ng/mL with precision of 7.4%.

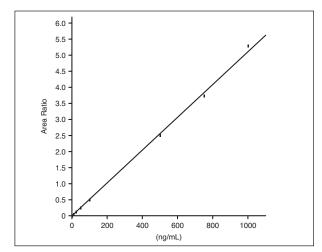


Figure 3. Linear dynamic range of 1 to 1000 ng/mL with an $\rm r^2$ of 0.9981 for cortisone standards

| Compound | n | Average Endogenous Level (ng/mL) | Precision |
|----------------|---|-------------------------------------|-----------|
| Hydrocortisone | 6 | 17.3 | 11.1 % |
| Cortisone | 6 | 55.6 | 7.4 % |

Table 3. Summary of hydrocortisone and cortisone results after extraction using SOLA well plates

Corticosterone results

Corticosterone standards prepared in water were linear over the dynamic range of 1 to 1000 ng/mL with an r² of 0.9998 (Figure 4). QC samples were extracted in triplicate at both low and high concentrations of 15 ng/mL and 600 ng/mL. Precision for each QC level were less than three percent relative standard deviation (Table 4). Overspikes were run in duplicate at a concentration of 250 ng/mL and used to calculate the percentage recovery level for corticosterone of 131.2%. No carryover was observed for corticosterone (Table 5).

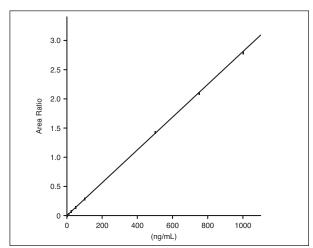


Figure 4. Linear dynamic range of 1 to 1000 ng/mL with an $\rm r^2$ of 0.9998 for corticosterone standards

11-α hydroxyprogesterone results

11-α hydroxyprogesterone standards prepared in water were linear over the dynamic range of 1 to 1000 ng/mL with an $\rm r^2$ of 0.9992 (Figure 5). QC samples were extracted in triplicate at both low and high concentrations of 15 ng/mL and 600 ng/mL. Precision for each QC level were less than three percent relative standard deviation (Table 4). Overspikes were run in duplicate at a concentration of 250 ng/mL and used to calculate the percentage recovery level for

11-alpha hydroxyprogesterone of 130.8%. No carryover was observed for 11- α hydroxyprogesterone (Table 5).

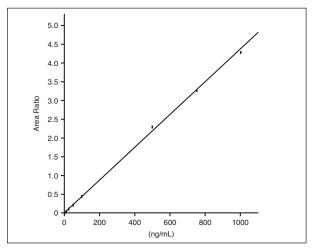


Figure 5. Linear dynamic range of 1 to 1000 ng/mL with an $\rm r^2$ of 0.9992 for 11- α hydroxyprogesterone standards

| Compound | n | QC Level | Accuracy | Precision |
|--------------------------|---|----------|----------|-----------|
| Corticosterone | 6 | High | -2.52 % | 1.3 % |
| Conticosterone | 6 | Low | -1.81 % | 2.5 % |
| 11-α hydroxyprogesterone | 6 | High | -3.83 % | 2.0 % |
| | 6 | Low | 1.50 % | 3.4 % |

Table 4. Summary of corticosterone and 11- $\!\alpha$ hydroxyprogesterone results

| Standard | Corticosterone | 11- $lpha$ hydroxyprogesterone |
|-----------------------------|----------------|--------------------------------|
| S1 response (peak area) | 1276 | 2490 |
| Total carryover (peak area) | 174 | 192 |
| % of S1 response | 13.6 | 7.7 |

Table 5. Summary of carryover results

Conclusion

SOLA SPE well plates and Accucore RP-MS can be successfully used to extract and quantify four steroids from urine. The results show good linearity, accuracy and precision for all compounds demonstrating the capability of both the SOLA well plate and the Accucore RP-MS column.

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