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Maintaining Sensitivity and Reproducibility with the JetClean Self-Cleaning IonSource for Pesticides in Food and Feed

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Introduction

The global agricultural industry uses over a thousand pesticides for food and foodstuffs cultivation. Producers are compelled to use pesticides to meet the growing demand for reasonably priced food, resulting in the need for pesticide residue monitoring in commodities worldwide. Concurrently, simple sample preparation practices, such as QuEChERS are routinely used for the preparation of food and feed samples, often leaving significant amount of matrix in the extracts. Analytical laboratories are challenged by these matrix residues, which with time negatively affects the response of the analyzed pesticides, and eventually requires source cleaning. Agilent's JetClean self-cleaning ion source (JetClean) reduces the need for manual source cleaning while still allowing for the analysis of complex samples without losing sensitivity and reproducibility.

The Agilent JetClean utilizes carefully monitored hydrogen gas (H_2) introduction to the source, controlled by Agilent's MassHunter Data Acquisition Software. The appropriate H_2 flow (in the μ L/min range) generates conditions that clean the surfaces of the source, the lenses and other components. These actions aid in maintaining a stable detection environment and provide for response stability of the pesticides in difficult matrices.

JetClean has two operational modes:

- 1. Acquire and Clean (or on-line) mode, when H₂ is running during the analysis
- 2. Clean only (or off-line) mode when H₂ is introduced only post run or post sequence

Experimental

Methodology: The analysis was conducted on an Agilent 7890B GC and 7010 Series Triple Quadrupole GC/MS. See Tables 1 - 3 for method parameters. The system was configured with a Multimode Inlet, equipped with an ultrainert liner (p/n: 5190-2293). The inlet was then connected to two HP-5ms UI columns (15 m \times 0.25 mm \times 0.25 µm; p/n: 19091S-431 UI) coupled to each other through a purged ultimate union (PUU) for the use of backflushing (see Figure 1).

The H₂ cleaning was operated in the "Acquire & Clean" mode which allowed constant H₂ flow during the analytical runs.



Figure 1. Column Configuration for Optimal MRM Application.

Timing	1.5 min during post-run		
Oven temperature	310 °C		
Aux EPC pressure	~50 psi		
Inletpressure	~2 psi		

Table 4. Matrix Selection and Sample Preparation Used for Optimal MRM	Application
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Category	Matrix	Sample Prep		
High Sugar	Organic Honey	5 g honey/5 mL water, EN salts, EN dSPE General		
		- 10400		

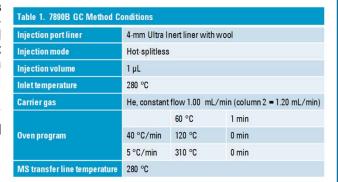


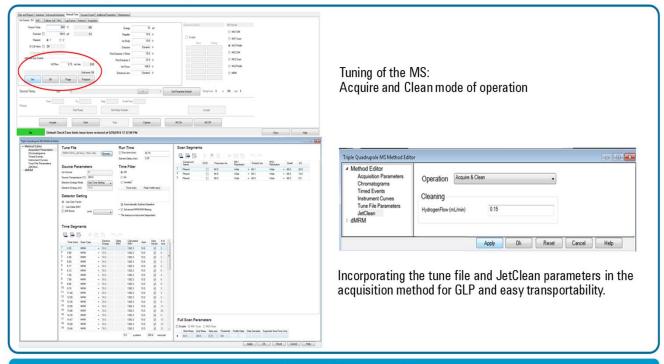
Table 3. 7010 MS/MS Parameters					
Electron Energy	70 eV				
Tune	atunes.eihs.tune.xml				
EM gain	10				
MS1 & MS2 resolution	Wide				
Collision Cell	1.5 mL/min N ₂ & 2.25 mL/min He				
Quant/Qual transitions	Matrix Optimized				
Dwell times	Time Segment (TS) specific*				
Source temperature	300 °C				
Quad temperatures	150 °C				
JetClean:	Acquire & Clean mode				

*The dwell times in each TS were the same, all with values over 10msec, resulting in a scan rate of ~5 scans/sec for the TS.



Experimental

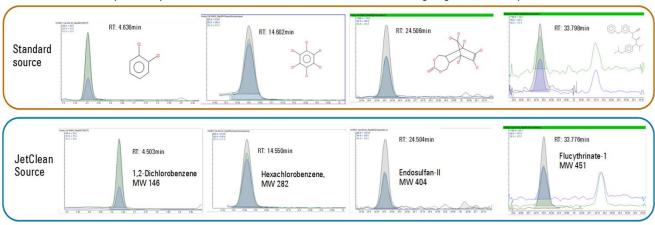
Operation: JetClean was initiated in the acquire and clean mode for this application. The MassHunter software allowed for the simple setup and operation of the process, all controlled in the MS domain.



Results and Discussion

Agilent's Self-cleaning ion source has been successfully used for the extended, 64 analyte PAH analysis in environmental and labs and also in food laboratories, resulting in remarkable precision, accuracy, linearity and detection levels, which were sustained for extended periods of time (many months) without manual cleaning. Considering the benefits of the Self-cleaning ion source for PAH analysis, the use of the JetClean applied for cleaning the MS source for pesticide analysis.

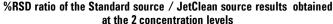
Chromatographic Performance: The following chromatograms show analytes at 2.5 pg concentrations, eluting at the beginning, middle and at the end of the chromatographic run. The ion plots are of target compounds and their respective matrix optimized MRM transitions in organic honey using standard source configuration and with the JetClean source. The JetClean benefits on peak shape and baseline are more obvious on the later eluting, higher MW analytes.

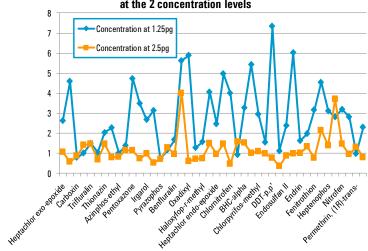


Results and Discussion

Quantitative results: Table 5 lists the R^2 values and the statistically derived MDLs for representative target analytes of the over 170 various pesticides tested. The calibration ranged from 0.12 pg/ul - 50pg/uL for the majority of the analytes, although some were not included at the lowest level. The resulting R^2 values were very comparable by both source type. The MDLs were calculated from 10 replicate measurement of 1.25 pg/µL concentration spiked honey extract using 99% confidence level. Lower MDLs were obtained for the majority of the analytes using the JetClean source, with an average of 0.151 pg MDL for the standard source and 0.081pg for the JetClean source. The replicate measurements performed at 1.25pg level resulted lower %RSD using the JetClean source, although they were comparable at the 2.5 pg level.

Table 5.						
	R ²		MDL (pg)			
Analyte	STD	JetClean	STD	JetClean		
Heptachlor exo-epoxide	0.994	0.992	0.085	0.022		
Endrin ketone	0.993	0.996	0.041	0.016		
Carboxin	0.991	0.994	0.021	0.025		
Profenofos	0.994	0.996	0.023	0.036		
Trifluralin	0.997	0.994	0.088	0.033		
Alachlor	0.996	0.978	0.119	0.050		
Thionazin	0.994	0.994	0.097	0.038		
Dimethoate	0.999	0.992	0.150	0.028		
Azinphos-ethyl	0.999	0.995	0.063	0.071		
Fenthion sulfone	0.996	0.994	0.046	0.040		
Pentoxazone	0.998	0.993	0.137	0.034		
Iprodione	0.993	0.999	0.041	0.029		
Irgarol	0.993	0.997	0.117	0.037		
Phosphamidon II	0.996	0.988	0.235	0.031		
Pyrazophos	0.994	0.996	0.042	0.056		
Terbufos	0.994	0.996	0.078	0.032		
Benfluralin	0.991	0.994	0.143	0.042		
Ethofenprox	0.996	0.992	0.161	0.036		
Oxadixyl	0.994	0.994	0.245	0.030		
Endosulfan I	0.995	0.996	0.027	0.032		
Haloxyfop-r-methyl	0.999	0.994	0.085	0.060		
Tetrachlorvinphos, E-isomer	0.998	0.996	0.112	0.029		
Heptachlor endo-epoxide	0.999	0.992	0.100	0.042		
Methoxychlor, p,p'-	0.994	0.996	0.170	0.046		
Chlornitrofen	0.996	0.997	0.070	0.020		
Bendiocarb	0.994	0.937	0.719	1.263		
BHC-alpha	0.995	0.995	0.221	0.098		
Chlorobenzilate	0.996	0.995	0.557	0.118		
Chlorpyrifos-methyl	0.994	0.993	0.270	0.075		
DBCP	0.995	0.986	0.155	0.208		
DDT-p,p'	0.993	0.996	0.427	0.105		
Dichlorobenzene, 1,2-	0.999	0.993	0.129	0.184		
Endosulfan II	0.991	0.998	0.037	0.040		
Endosulfan sulfate	0.997	0.996	0.136	0.036		
Endrin	0.992	0.994	0.045	0.025		
Ethoprophos	0.995	0.995	0.072	0.016		
Fenitrothion	0.993	0.991	0.313	0.060		
Flucythrinate I	0.997	0.992	0.040	0.023		
Heptenophos	0.992	0.996	0.264	0.045		
Hexachlorobenzene	0.999	0.997	0.227	0.116		
Nitrofen	0.990	0.995	0.097	0.030		
Parathion-methyl	0.994	0.995	0.272	0.070		
Permethrin, (1R)-trans-	0.997	0.968	0.033	0.105		
Pirimiphos-methyl	0.996	0.994	0.152	0.042		





Conclusions

Approximately 170 various pesticides were analyzed in organic honey on the 7010 Series Triple Quadrupole GC/MS using standard and JetClean source in the Acquire and Clean mode, utilizing carefully introduced hydrogen flow. The JetClean control is included in the MassHunter software with easy setup and operation.

The chromatographic peak shape and baseline was improved using the Jetclean source particularly for the late eluting compounds. The calibration resulted very comparable R^2 values by both source, while the MDLs obtained at 1.25 pg level resulted lower values using the JetClean source. The %RSDs were comparable at higher, 2.5pg/µl level. The results indicate that the JetClean source meets and exceed the performance delivered by the standard source.

Further study is undergoing to identify how the source maintenance period is extended when JetClean is applied compared to the standard source.

Reference

¹Anderson, Kim A., et al. "Modified ion source triple quadrupole mass spectrometer gas chromatograph for polycyclic aromatic hydrocarbon analyses." *Journal of Chromatography A* 1419 (2015): 89-98.